

Cuticular microstructures of abdominal tergites and sternites of *Cloeon* sp. (Ephemeroptera; Baetidae) during post-embryonic development

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The postembryonic changes of the external microstructure of the cuticle on the abdominal tergites and sternites of *Cloeon* sp. (Ephemeroptera:Baetidae) are described. An inventory of the structures as the aquatic nymph grows and becomes transformed into a terrestrial subimago and then an imago, is provided. It is found that sensillar diversity on nymphal tergites increases with development, but remains unchanged on the sternites. While nymphs and subimago possess various types of *sensilla trichoidea*, *sensilla basiconica* are apparent on the imaginal abdomen only. Furthermore, the scaled and spiny cuticle of the nymph becomes relatively smooth in the adult stages. The distinct changes which structure and distribution of the scales as well as marginal spines on the abdominal tergites and sternites of the nymphs undergo during development, must find a reflection in behavioural changes of the larvae. Finally, the taxonomic implications of the observed changes are discussed.

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1. Introduction

Mayflies represent an important component of the ecology of many boreal as well as high altitude freshwater systems, as their nymphs, being mostly primary consumers, are actively involved in the processing and cycling of nutrients (Cummins

1973; Merritt et al. 1984). They are also unique among the insects in having a subimago - instead of a pupa - following the nymphal stage in their life cycle.

The microstructures on the cuticle, especially that of the nymphal abdominal tergites of many ephemeropterans have been used for defining taxa

as well as for constructing phylogenetic relationships (Müller-Liebenau 1973, 1983, 1984, 1985; Tsui & Peters, 1975) and scanning electron micrographs of nymphal spines and setae have accompanied the description of a new genus of Leptophlebiidae (Pescador 1997). Yet, no systematic study is available on the postembryonic development of these structures in the different developmental stages of the nymphs of any mayfly species. Furthermore, practically nothing is known about the changes in the external microstructure of the cuticle as the aquatic nymph is transformed, first to a terrestrial but reproductively immature subimago, and finally to a reproducing imago.

The present paper describes the external microstructure of the abdominal cuticle of different developmental stages of the nymph, and those of the subimago and imago as well, of one species of *Cloeon* (Ephemeroptera: Baetidae) from Shillong, Meghalaya State, India, by scanning electron microscopy. Besides providing an inventory of cuticular microstructures in this species, which may subsequently be compared with those of other taxa (*cf.* recently published descriptions of larval sensilla on thorax and abdomen of Saltatoria, Mantodea, Blattaria, Heteroptera, Lepidoptera, and Diptera: Green & Hartenstein 1997), the findings are of relevance also to nymphal and adult behaviour.

2. Material and methods

Nymphs of one species of *Cloeon* sp. were collected from Ward Lake, a small artificial lake in Shillong (25°34'N; 91°52'E), Meghalaya State, India, and sorted into 4 developmental stages on the basis of wingpad development (Clifford 1970). Despite considerable effort, identification to species level was not possible, but this fact must not distract from the goal of the investigation for which the knowledge that throughout the investigation only one species was used was the most critical factor. According to Müller-Liebenau (pers. comm.) the species is different from the common Indian species *Cloeon marginalis*.

Nymphs were reared in the laboratory to obtain subimagines and imagines. The abdomens of 20 nymphs, subimagines, and imagines (10 each for tergites and sternites), were fixed for 2 h in 2-5% glutaraldehyde and postfixed for 2 h in 2% osmium tetroxide, both fixatives buffered with 0.1M cacodylate to a pH of 7.2. These were dehydrated in a graded series of acetone, dried in a critical point dryer, mounted on aluminium stubs, and coated with gold with the

aid of a fine coat ion-sputter JFC 1100. The specimens were examined and photographed in a JSM 35 CF scanning electron microscope. Sensillar dimensions were obtained from photographs and averages with standard deviations are based on 20 measurements for each type of sensilla. To establish the porosity of sensilla, the entire subimago and imago were immersed for 2 days in 70% ethanol containing 0.1% silver nitrate and examined in a Leitz-Ortholux phase contrast microscope (Nayak & Singh 1983).

3. Results

3.1. Cuticular microstructures on abdominal tergites

Data on the lengths, socket diameters, and other morphometric details of the different cuticular microstructures in the various nymphal stages are presented in Table 1.

3.1.1. Stage 1 nymph

The surface of every tergal segment is beset with numerous scale-like structures with spiny margins (Fig. 1). Socketed sensory hairs, designated as *sensilla trichoidea* type 1, are strewn among the scales (Fig. 1). The lateral regions and the posterior extremities of tergites are provided with several *sensilla campaniformia* in the form of roughly circular to oblong cuticular domes (Fig. 2). Spines on the hind margins of tergal segments are very poorly developed (Fig. 2).

3.1.2. Stage 2 nymph

Some of the spiny scales of stage 1 apparently develop into tongue-shaped cuticular plates with serrated margins (Fig. 3). This change is more pronounced on the lateral regions of the tergites. *Sensilla trichoidea* type 1 and *sensilla campaniformia*, similar to those in stage 1, are also present. Additionally, *sensilla basiconica*, arranged in 3 clusters, one in the anteromedial region, and one each in the anterolateral regions of the tergites, are noticed also in this stage. Each cluster has 8-10 short, socketed sensilla with curved and tapered, but somewhat blunt, tips (Fig. 4). Spines on the hind margin are distinctly developed in

Table 1. Morphometric data of the cuticular structures on the abdominal tergites and sternites of *Cloeon* sp.

Cuticular structure	Development stage	Length in μm (\pm s.d.)	Diam./width in μm (\pm s.d.)
Nymph			
1. <i>Sensilla trichoidea</i>			
(i) type 1	1-4	17.1 \pm 7.8	1.7 \pm 0.3 a
(ii) type 2	2-4	30.1 \pm 4.2	1.5 \pm 0.3 a
2. <i>S. basiconica</i>			
3. <i>S. campaniformia</i>			
(i) tergites	1-4	n.a.	4.8 \pm 0.5 b
(ii) sternites	1-4	n.a.	3.6 \pm 0.5 b
4. Tongue-shaped plates			
5. Hind margins spines			
(i) tergites			
	1	2.5 \pm 0.8	0.41 \pm 0.16 c
	2	9.2 \pm 3.9	3.8 \pm 1.6 c
	3-4	21.1 \pm 4.8	7.2 \pm 2.8 c
(ii) sternites			
	1	1.9 \pm 0.6	2.4 \pm 0.8 c
	2	9.0 \pm 3.7	5.3 \pm 1.7
	3-4	15.2 \pm 4.4	5.8 \pm 1.5 c
Subimago			
1. Hair sensilla			
2. <i>S. trichoidea</i>			
3. <i>S.</i> with bulbous base			
	female	8.9 \pm 2.5	3.9 \pm 0.4 c
Adult			
1. <i>S. trichoidea</i>			
2. <i>S. basiconica</i>			
3. Spinules on sternites			
		7.6 \pm 0.8	0.87 \pm 0.1 a
		6.1 \pm 1.0	3.4 \pm 0.7a
		3.7 \pm 0.5	1.5 \pm 0.2 c

a = socket diameter; b = diameter; c = basal width; n.a. = not applicable

segments 7-9, and comprise shorter spines interspersed among relatively longer ones (Fig. 5).

3.1.3. Stage 3 nymph

The "tongue-shaped" plates that were mainly confined to the lateral regions of the tergites in the stage 2 nymph, are uniformly distributed throughout their surfaces in this stage. In addition to *sensilla trichoidea* type I and *sensilla basiconica*, *sensilla trichoidea* type 2, having a narrow base followed by a relatively broad and flattened area, are also present. These sensilla lack well-defined sockets at their bases (Fig. 6). *Sensilla campaniformia*, as in the previous stages, are also found. Spine formation on the hind margins of tergites

commences from segment 2 (Fig. 7), although these are well-developed in segments 5-9.

3.1.4. Stage 4 nymph

Cuticular microstructures on the tergite surface are similar to those found in stage 3. Hind margin spines are, however, well-developed from segment 2 (Fig. 8).

3.1.5. Subimago

The subimaginal abdominal tergite surface is markedly different from that of the nymphs. All the tergites are covered with unsocketed hairs,

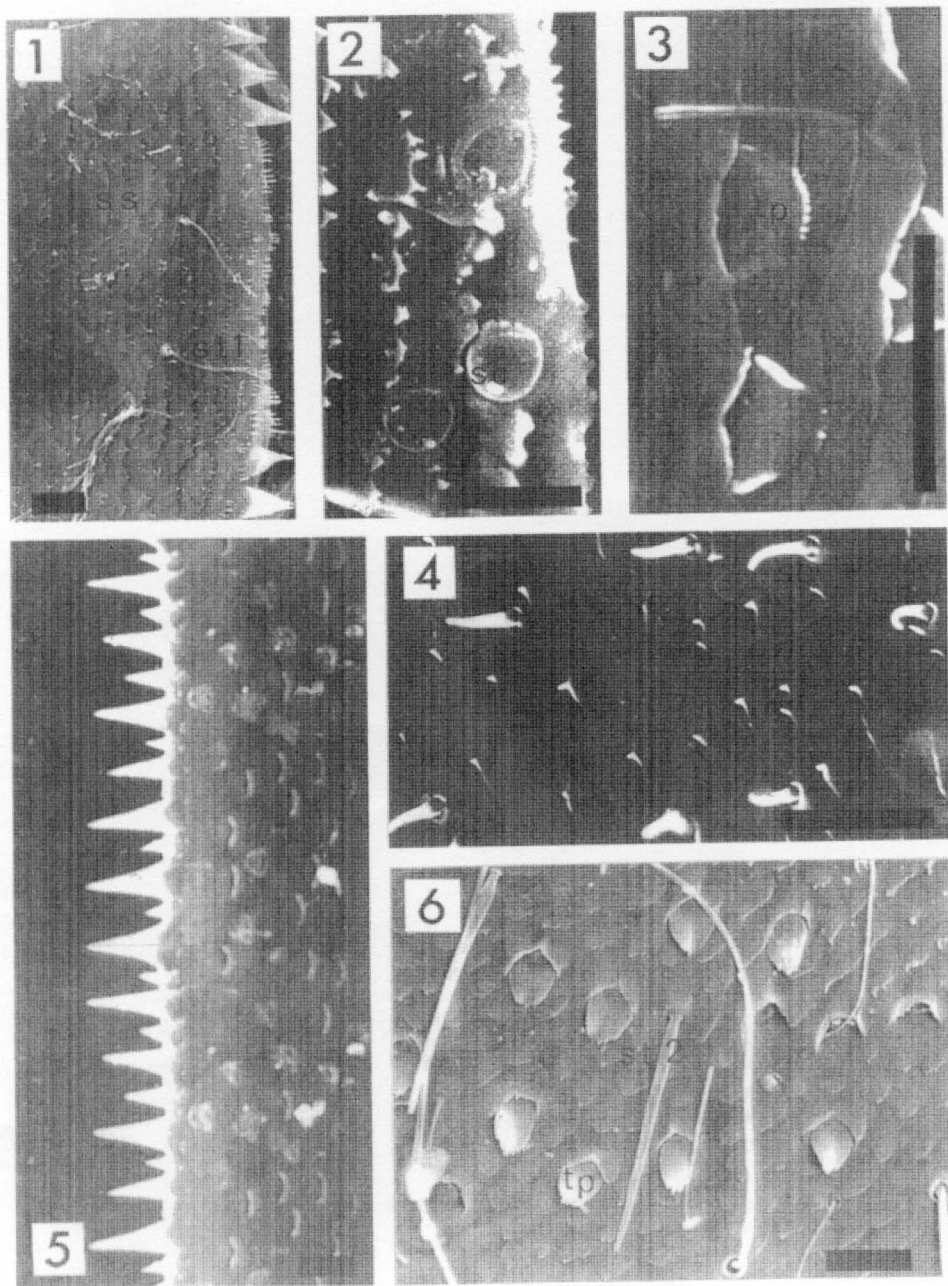
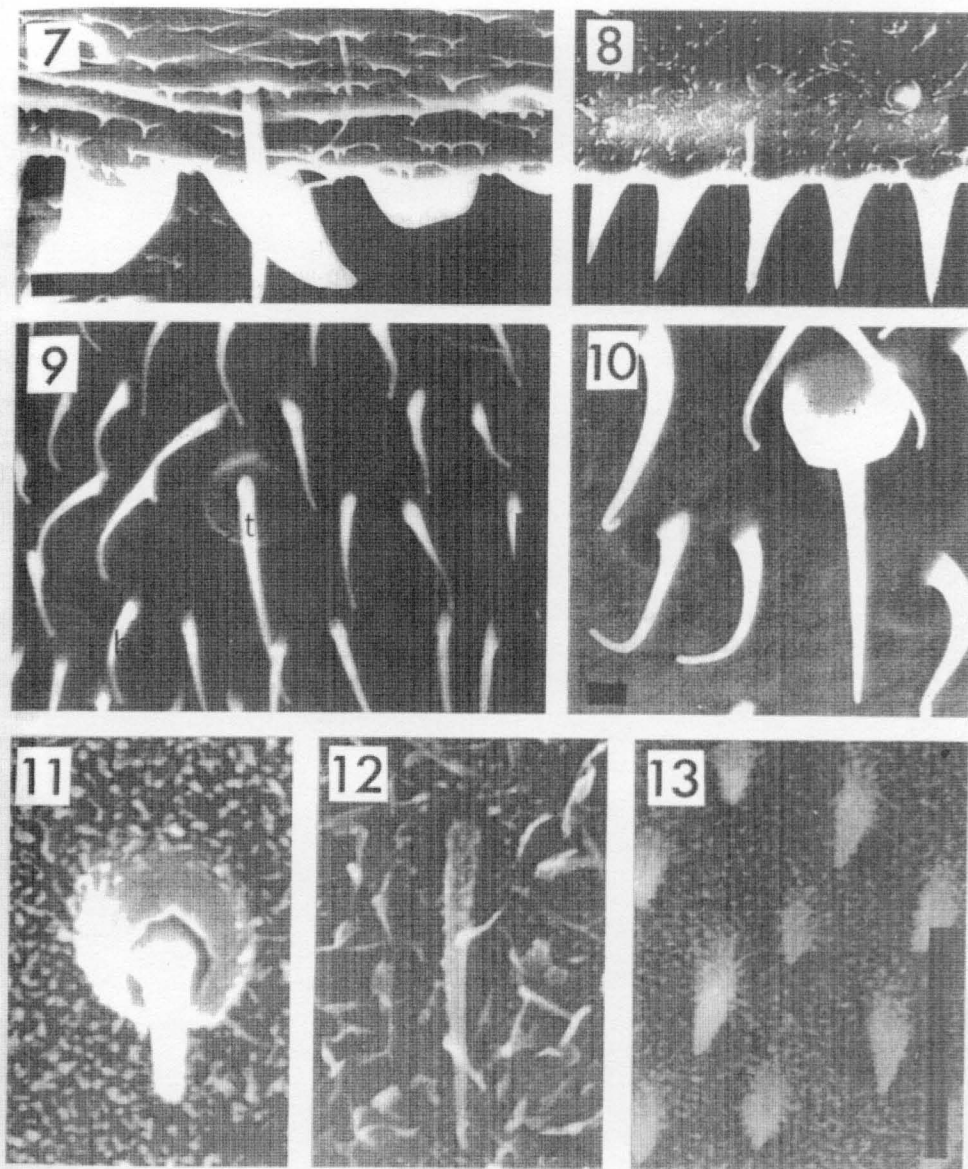


Fig.s 1-6. 1. Surface of abdominal tergite 9, stage 1 nymph; ss = spiny scales; st1 = *sensilla trichoidea* type 1. Bar: 10 μ m; 2. Hind margin of tergite 7, stage 1 nymph; sc = *sensilla campaniformia*. Bar: 10 μ m; 3. Surface of tergite 5, stage 2 nymph; tp = tongue-shaped plate. Bar: 10 μ m; 4. Lateral region, tergite 5, stage 2 nymph; sb = *sensilla basiconica*. Bar: 10 μ m; 5. Hind margin of tergum 7, stage 2 nymph. Bar: 10 μ m; 6. Surface of tergite 6, stage 3 nymph; st2 = *sensilla trichoidea* type 2; tp = tongue-shaped plate. Bar: 10 μ m.



Figs. 7-13. 7. Hind margin of tergite 2, stage 3 nymph. Bar: 10 μ m; 8. Hind margin of tergite 2, stage 4 nymph. Bar: 10 μ m; 9. Tergite surface, subimago; hs = hair sensilla; st = *sensilla trichoidea*. Bar: 10 μ m; 10. Tergite surface, female subimago; bs = sensilla with bulbous base. Bar: 1 μ m; 11. *Sensilla basiconica* on tergite of imago. Bar: 1 μ m; 12. *Sensilla trichoidea* on tergite of imago. Bar: 1 μ m; 13. Spines on sternite of imago. Bar: 10 μ m.

largely oriented along the long axis of the body, their tips facing posteriorly. They gradually taper from their bases, and have curved and slightly bulbous tips (Fig. 9). Scattered among these hairs are a few *sensilla trichoidea* emerging from circular sockets (Fig. 9). Furthermore, only the female subimago possesses clusters of sensilla in the anterolateral and anteromedial regions of its tergites. Individual clusters have 10-11 sensilla, each with a prominent bulbous unsocketed base with a spine-like projection (Fig. 10). The hind margin of the subimaginal tergites, in contrast to the nymphs, are devoid of any spine or projection.

3.1.6. Imago

The surface of imaginal tergites shows extremely minute cuticular foldings. The dense hairs of the subimago are absent. However, each imaginal tergite has 3 distinct clusters of *sensilla basiconica* in the anterior region, each cluster having 10-11 basiconic pegs. The location of these clusters as well as the orientation of their constituent sensilla appear to be fairly constant. While the sensory pegs in the central cluster point downward and backward, those in the lateral clusters point laterally. These pegs emerge from circular sockets, and taper gradually (Fig. 11). Furthermore, *sensilla trichoidea* that emerge from circular punctures in the cuticle are sparsely and irregularly distributed on the surface of all tergal segments (Fig. 12). The cuticular foldings mentioned earlier are more pronounced around the bases of these sensilla. The hind margins of the tergites are smooth, as in the subimago.

Morphometric data on the cuticular structures of the subimaginal and imaginal tergites are provided in Table 1.

3.2. Cuticular microstructures on the abdominal sternit

3.2.1. Nymph

As in the case of tergites, the spiny scales in the stage 1 nymph develop into "tongue-shaped"

plates with serrated margins in stage 2 and 3 nymphs. However, unlike the tergites, the sternites possess *sensilla trichoidea* types 1 and 2, as well as the basiconic pegs in stage 1 itself. *Sensilla campaniformia*, somewhat smaller than those found on the tergites (Table 1) are also present. The spines on the hind margin are present from the 8th segment onwards in stage 1, from the 7th in stage 2, from the 6th in stage 3, and from the 3rd in stage 4 nymphs.

3.2.2. Subimago

These are similar to the tergites, barring the fact that sensilla found only on the female tergites are absent from the sternites.

3.2.3. Imago

Sensilla similar to those on the tergites can be seen, although the sternites are beset with numerous small spines that are prominent in their posterolateral regions (Fig. 13). Data on their lengths and widths are provided in Table 1.

3.3. Species Characteristics

Some taxonomically important characters such as male genitalia, female forewing, mature nymphal mouthparts, etc. are shown in the line drawings of Fig. 14.

4. Discussion

Obviously, those structures on the abdominal tergites and sternites of nymphs and adults of *Cloeon* sp. identified as 'sensory' in nature have a role to play in the aquatic phase of the species. Their exact function(s), however, cannot be ascertained until results are available on their innervation and other ultrastructural attributes. Nevertheless, extrapolations from SEM and TEM-based studies on insect sensilla generally enable us to suggest some of their possible functions.

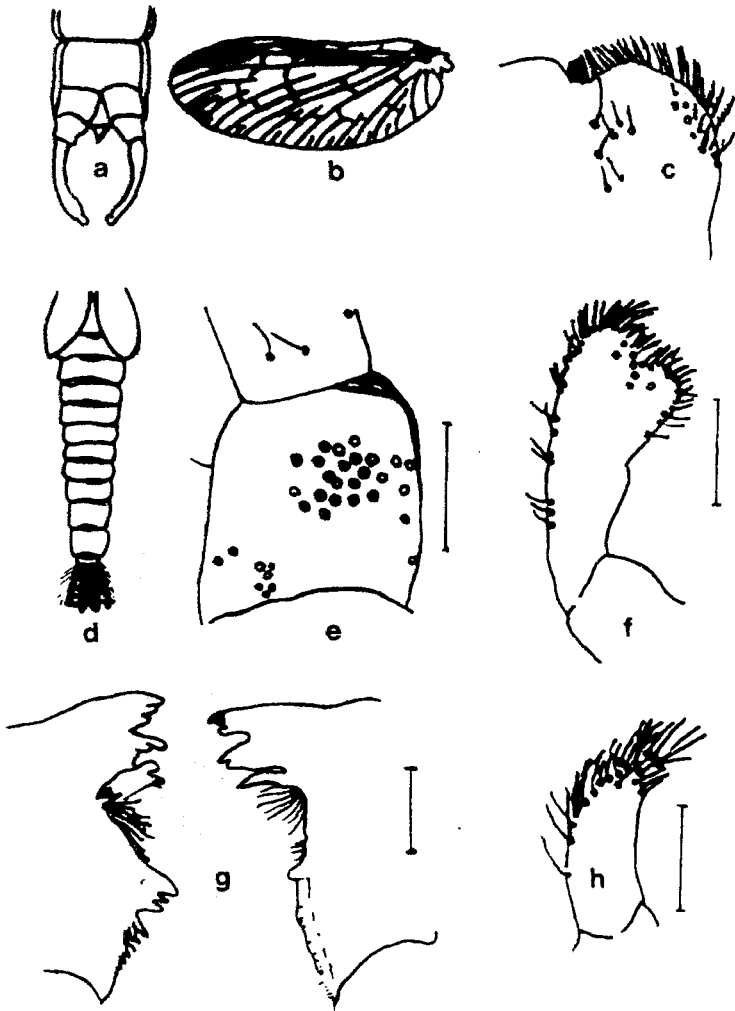


Fig. 14. Taxonomically important characteristics of the *Cloeon* sp. used in this investigation. — (a) Genitalia of male imago. — (b) forewing of female imago. — (c) labrum of mature nymph. — (d) nymphal exuvium. — (e) basal antennal segment of mature nymph. — (f) tip of labial palp of mature nymph. — (g) mandibles of mature nymph. — (h) tip of maxillary palp of mature nymph. The scale is 0.1 mm.

Sensilla trichoidea type 1, with their slender, pliable setae, and sockets that allow movement in all directions, are likely to be mechano- or contact chemosensilla (Frazier 1985, Crouau & Crouau-Roy 1991) and may also be used for cleaning detrital and other debris from the abdominal surface as well (Honegger 1977, Zack & Bacon 1981). *Sensilla trichoidea* type 2, in the absence

of sockets, may only serve simple tactile functions (Schmidt & Smith 1987). The basiconic pegs, on the other hand, are likely to be chemoreceptors. The nymphs of all stages have *sensilla campaniformia* in the lateral regions and along the posterior margins of their tergites and sternites. The proprioceptive functions of these sensilla are well known (Zill & Moran 1981a, 1981b; Spinola

& Chapman 1975, Schmidt & Smith 1987).

The present study reveals that the sensillar diversity on the tergites increases from stage 1 to stage 3 and 4 nymphs, which suggests that the tergites play a progressively greater role in monitoring the environment as nymphal development proceeds. In contrast, all the 3 types of sensilla are present on the stage 1 sternites, which, thus, are likely to start performing diverse sensory functions early in nymphal life. In damselfly larvae Richardson and Anholt (1995) observed significant changes in 11 behavioural patterns as the larvae grew older: some (e.g. abdomen-raise, side-crawl) decreased while others (e.g. 180°-turns, feeding, abdomen wave) increased. On the basis of our findings, one would expect to find similar (but not necessarily the same) behavioural changes in maturing mayfly nymphs.

The spiny plates on the nymphal abdomen, along with the spines on the hind margins of tergites, have been used for delimiting taxa (Müller-Liebenau 1973, 1983, 1984, 1985) and featured in the description of the new leptophlebiid genus *Gonserrelus* (Pescador 1997). However, nothing precise is known about their functional significance. In the marine isopod *Glyptonotus antarcticus* microscales on the cuticle have been shown to be effective in discouraging colonization by foraminiferans and other epizoots (Meyer-Rochow 1980) and Peckarsky (1987) observed that an abdomen-bending "scorpion"-like posture by *Ephemerella* nymphs resulted in the cerci and the flexed abdominal spines acting as a defence mechanism against predatory stoneflies. The spiny scales of *Cloeon* could, therefore, serve to discourage bacterial, fungal, and protozoan infestations, and together with the marginal spines may facilitate defence against predators as well.

The spiny scales or plates, however, do not remain constant throughout the larval life, but exhibit certain changes during nymphal development. While they are in the form of somewhat indistinct folds of the cuticle with spiny edges in stage 1, they get transformed into distinct "tongue-shaped" spiny plates in the lateral regions of the abdomen in stage 2, and subsequently spread to the other areas of the abdomen in stages 3 and 4. Similarly, the marginal spines first develop on the distal abdominal segments, and as development proceeds, they gradually appear on the proximal

segments as well. This observation implies that only the mature 4th-stage nymph has to be used for taxonomic descriptions and comparisons, as nymphs belonging to the same species (but to different developmental stages), will show differences in the anatomical features of the surface and hind margins of their tergites, leading to possible mistakes in identifications.

The nymphal scales disappear in the subimago, whose entire body, including the abdomen, is covered with a dense coating of unsocketed hairs. Besides serving tactile functions, these hairs probably act as a hydrofuge device to prevent drowning of the subimago as it emerges on the water surface, and subsequently to provide heat insulation as terrestrial existence commences. Similar roles for cuticular hairs in general, have been suggested by Neville (1975). The socketed trichoid sensilla found on the subimaginal abdomen may serve either a mechano- or a chemoreceptive function, while the other type with a bulbous base is found only in the female, and is an expression of sexual dimorphism. Staining with silver nitrate reveals that the base is perforated, thereby indicating its chemosensory nature.

The numerous hairs of the subimago are discarded during imaginal moult, leaving the surface smooth. Lameere (1917) suggested that this transformation facilitated flying by reducing the body weight, but it also diminishes friction against air. It has been shown that mayflies that do not shed their subimaginal skin do not have sustained mating flights (Needham et al. 1935). The imaginal tergites and sternites, however, retain *sensilla trichoidea*, likely to be mechanoreceptors, and *sensilla basiconica* that may be involved in olfactory recognition of mates during swarming. It is known that mayfly eyes, including the highly developed dorsal turbinate eyes (which are present in the male imago, but not sub-imago: Meyer-Rochow 1971) of the Baetidae, to which *Cloeon* sp. belongs, are not designed for distance perception of small flying objects (Wehner 1981, Wehner & Srinivasan 1984). This drawback was overcome by the evolution of swarming, which facilitated detection of females by males as the former enter the swarm and can be recognized at a closer distance (Fischer 1991). Critical for recognition by sight, however, is that environmental light conditions permit the visual signals to be registered and

understood. Whether in the absence of light sexual recognition takes place or blinded individuals are able to mate is unknown, but it does seem feasible that the basiconic sensilla could be part of a 'back-up system' aiding the two sexes to detect and locate each other in conditions of poor visibility. Following pair formation, the minute spines on the sternites most probably enable better grip during mating.

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